

Abstract

Precision cancer immunotherapy targeting mutations expressed by cancer cells have proven to effectively control the tumor of patients in multiple clinical trials [1,2]. However, the selection of immunogenic T cell neo-epitopes remains challenging and many epitopes selected using traditional methodologies fail to induce effector T cell responses. Poor performance may partially be due to inclusion of mutated epitopes cross-conserved with self-epitopes recognized by regulatory (Treg), anergic, or deleted T cells. Vaccination with self-epitopes can lead to weak effector responses, active immune suppression, and toxicity due to immune-mediated adverse effects. In addition, most cancer vaccine studies focus on the selection of CD8 T cell neo-epitopes due to an apparent lack of robust and accurate CD4 T cell epitope prediction tools.

We have developed Ancer, an advanced cancer T cell epitope identification and characterization tool, that streamlines the selection of both CD4 and CD8 T cell neo-epitopes from Next Generation Sequencing data. Ancer leverages EpiMatrix and JanusMatrix, state-of-the-art predictive algorithms that have been extensively validated in prospective vaccine studies for infectious diseases [3,4]. Distinctive features of Ancer are its ability to accurately predict Class II HLA ligands, or CD4 epitopes, with EpiMatrix, and to identify tolerated or Treg epitopes with JanusMatrix. In addition, screening candidate sequences with JanusMatrix enables to the removal of neo-epitopes that may trigger off-target events, which have in some cases abruptly halted the development of promising cancer therapies.

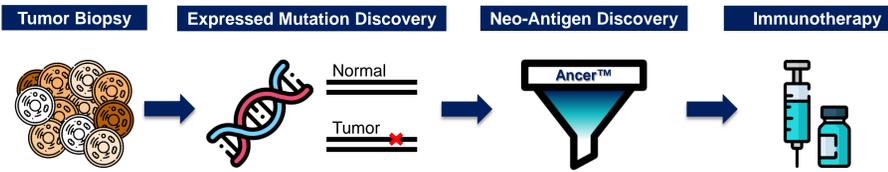
The performance of Ancer was assessed by retrospectively analyzing multiple published datasets. First, we evaluated HLA-Class-I-bound peptides detected by mass spectrometry and published by Abelin et al., Immunity 2017 [5]. Our analysis showed that 95% of eluted peptides were correctly predicted by EpiMatrix to bind to their respective HLA, while only 88% of these sequences were accurately predicted by NetMHCpan. In addition, the majority of eluted peptides were specifically identified by EpiMatrix as strong HLA ligands.

Second, we performed a retrospective analysis of a cancer immunogenicity study published by Strønen et al., Science 2016 where HLA A2-restricted neo-epitopes were validated in T cell assays [6]. Immunogenic sequences had significantly higher binding potentials, as estimated by EpiMatrix, compared to non-immunogenic sequences (p=0.0020). In contrast, no significant differences in predicted binding affinities were observed between these two subsets of sequences using public in silico prediction tools (p=0.0561).

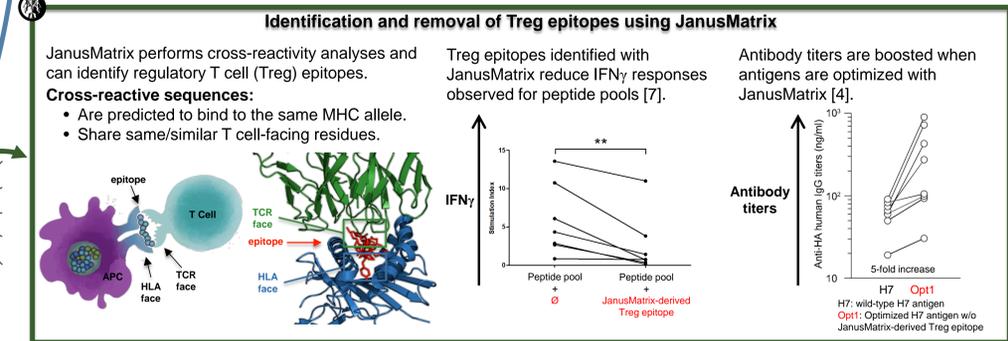
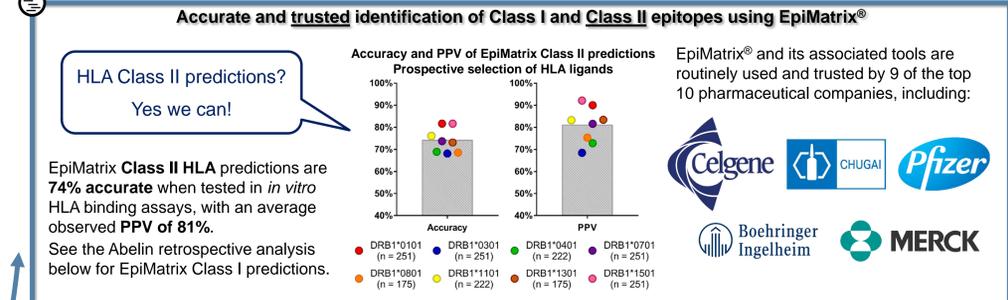
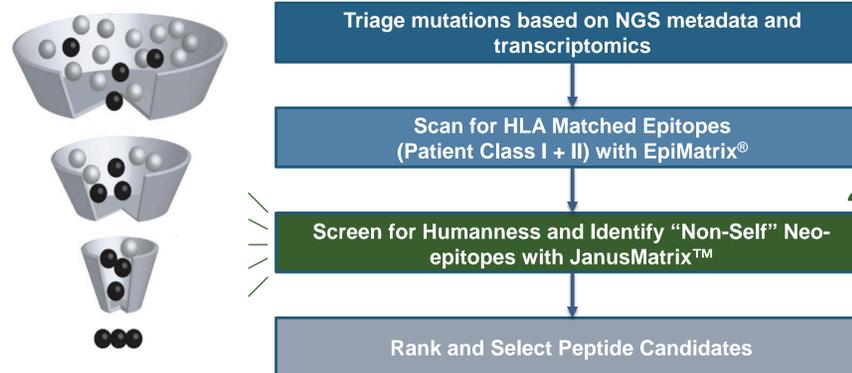
Lastly, Ancer divided immunogenic and non-immunogenic sequences from Strønen et al. with 72% accuracy outperforming the results obtained with public tools (21% accuracy) or with lengthy and costly in vitro characterization techniques (65% accuracy).

These results suggest that higher quality candidate targets are retrieved by Ancer, as compared to other conventional algorithms. CD4 and CD8 neo-epitopes with low Treg activation potential may then be used to support the development of safer and more effective personalized cancer vaccines. Future steps include the design of prospective studies to test the efficacy of Ancer-derived vaccines in the CT26 and GL261 murine cancer models.

Mutanome-Directed Cancer Immunotherapy Based on 20 Years of Experience in Epitope Mapping



Ancer™ Platform: the “Answer” to Cancer



Retrospective Immunoinformatic Analyses of Published Cancer Datasets

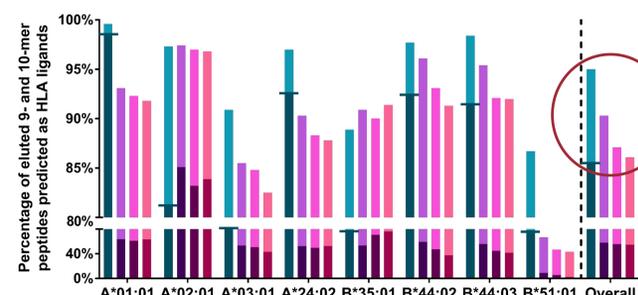
EpiMatrix® analysis of elution datasets from Abelin et al. Immunity 2017

- Summary of the Abelin et al. study [5]: over 26,000 peptides were eluted across 16 HLA-A or HLA-B monallelic cell lines.
- 6,284 9-mer and 2,301 10-mer HLA ligands from the Abelin et al. dataset were extracted for eight common HLAs (worldwide frequency over 5%) and scored with EpiMatrix® and NetMHCpan [8-10].
- 95% of eluted 9- and 10-mers were predicted to bind to HLA according to EpiMatrix® (standard Z-score cutoff of 1.64), while only ~88% of ligands were accurately recalled by NetMHCpan (500nM cutoff).

Ancer™ Analysis of Mutated Peptides from Strønen et al. Science 2016

- Summary of the Strønen et al. study [6]: neo-epitopes were identified with NetMHC and NetMHCpan but few peptides (21%) were immunogenic. *In vitro* peptide-MHC off-rates separate immunogenic from non-immunogenic peptides with 65% accuracy.
- Immunogenic peptides have greater binding potentials than non-immunogenic peptides, as measured by EpiMatrix®. No significant difference is observed with public *in silico* tools.

Head-to-head comparison of epitope prediction tools Common HLAs (worldwide frequency >5%)

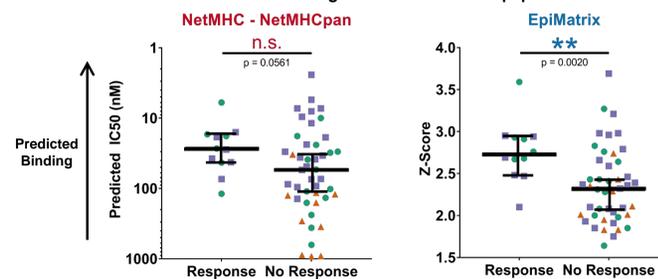


High accuracy of EpiMatrix Class I predictions compared to publicly available prediction tools.

Motifs not captured by NetMHCpan, but already accounted by EpiMatrix after its latest 2015 update?

More candidates are retrieved with EpiMatrix

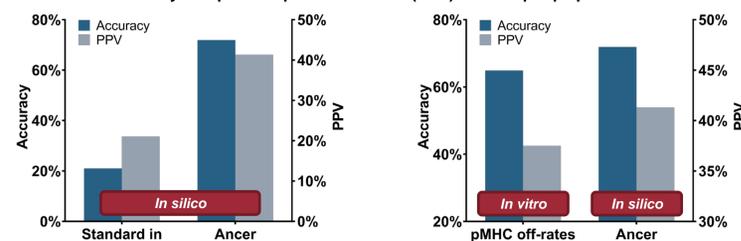
Predicted binding affinities of mutated peptides



Prediction of neo-epitope response with Ancer™ is improved over *in vitro* methods

- Ancer™, which uses EpiMatrix® and JanusMatrix™, can differentiate immunogenic and non-immunogenic peptides with 72% accuracy. PPV of Ancer™ is twice that of standard *in silico* tools.

Accuracy and positive predictive value (PPV) of neo-epitope predictors

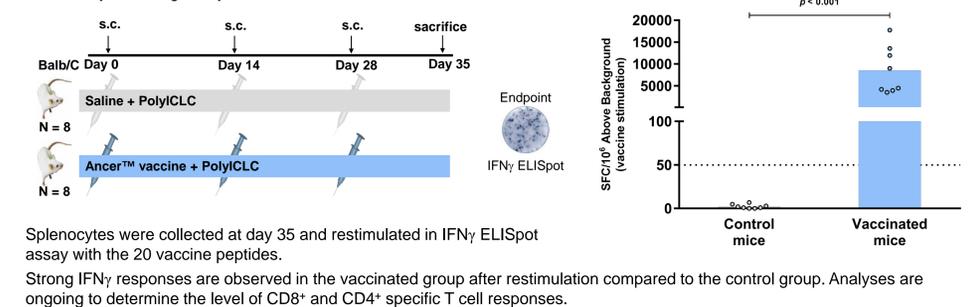


Ancer™ Prospective Studies

Ancer™ is central to the design of a new prospective murine studies using the CT26 syngeneic mouse model.



Preliminary immunogenicity results:



Conclusions

- Analysis of the MHC- and TCR-facing residues of T cell epitopes by JanusMatrix™ enables prediction of epitope phenotype.
- EpiVax's immunogenicity screening tools (EpiMatrix® and JanusMatrix™) are integrated into the Ancer™ platform for streamlined designs of personalized cancer vaccines.
- Sharper definition of neo-epitopes by immunoinformatic analyses may improve epitope selection for mutanome-directed cancer immunotherapy.
- Ancer™-derived vaccines are currently being evaluated in prospective studies using the CT26 syngeneic mouse model.

References

- Sahin U. et al., Nature. 2017 Jul 13;547(7662):222-226
- Ott P. et al., Nature. 2017 Jul 13;547(7662):217-221
- Moise L. et al., Hum Vaccin Immunother. 2015;11(9):2312-21.
- Wada Y. et al., Sci Rep. 2017 Apr 28;7(1):1283
- Abelin J. et al., Immunity. 2017 46, 315–326
- Strønen E. et al., Science. 2016; 352(6291), 1337-41.
- Liu R. et al., Hum Vaccin Immunother. 2015 11:9, 2241-2252
- Jurtz V. et al., J Immunol. 2017 Nov 1;199(9):3360-3368
- Nielsen M and Andreatta M, Genome Medicine 2016 8:33
- Hoof I. et al., Immunogenetics. 2009 61(1):1-13

Acknowledgments

We thank our colleagues at EpiVax for contributions to this work.

Some icons used in this poster were made by Freepik from www.flaticon.com and are licensed by CC 3.0 BY.